



HE002

The Association Analysis of Bisphenol A (BPA)-Responsive Genes and Dysregulated Genes  
in Autism Spectrum Disorder

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**Abstract**

Autism spectrum disorder (ASD) is a group of neurodevelopmental disorder characterized by deficits in 2 symptoms which are social and communication impairment and repetitive behaviors together with limited interests with the prevalence of 1 in 59 children in the United States. The exact cause of this disorder is still unclear but there is accumulating evidence that the susceptibility of ASD is influenced by the combination of multiple genes and environmental factors. Bisphenol A (BPA) is an endocrine disrupting chemical that has been associated with ASD. Recent studies have reported that children with ASD have increased BPA levels in the blood and the urine. However, it is still unclear whether BPA can alter the expression of genes associated with ASD. In this study, we conducted a series of bioinformatic analyses using CU-DREAMx program and pathway analysis to predict the association of BPA-responsive genes and ASD, and also predict biological functions impacted by BPA exposure. We found that BPA-responsive genes (i.e. *SOD1*, *MEF2C*, and *GNAS*) are significantly associated with ASD and ASD-related neurological functions. These genes may serve as good candidates for studying the effect of BPA exposure at the molecular level and its role in ASD susceptibility in the future.

**Keywords:** autism spectrum disorder, bisphenol A, endocrine-disrupting chemical, bioinformatics, CU-DREAMx

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## 1. Introduction

Autism spectrum disorder (ASD) is an early-onset neurodevelopmental disorder with abnormalities in two main functions: impairment in social interaction and communication, and repetitive behaviors and/or restricted interests. The prevalence of ASD recently reported by The Centers for Disease Control and Prevention (CDC) is 1 in 59 children in the United States (Baio, Wiggins et al. 2018). Although there is supporting evidence that some of the ASD cases may be related to genetic factor, the majority of ASD cases are idiopathic with unidentified genetic cause. The risk of ASD is thought to be influenced by gene-environment interactions including epigenetic mechanisms, including DNA methylation (Nguyen, Rauch et al. 2010, Moosa, Shu et al. 2018, Saeliw, Tangsuwansri et al. 2018) and histone modifications (LaSalle 2013, Sun, Poschmann et al. 2016), and exposure to environmental chemicals. Recent studies have revealed that ASD risk is associated with certain environmental pollutants, including endocrine-disrupting chemicals (EDCs) (Miodovnik, Engel et al. 2011), lead (Adams, Audhya et al. 2013), mercury (Geier, Audhya et al. 2010), and pesticides (Roberts, English et al. 2007). EDCs are a group of chemicals widely used in many

plastic products. The structure of such chemicals are similar to steroid hormones including estrogen, androgen, and thyroid hormone. BPA is the one of the EDCs mostly used in polycarbonate plastics and thought to disrupt hormone regulatory system in animals and humans. Once ingested, BPA passes through placenta and blood-brain barrier (Sun, Nakashima et al. 2002, Nishikawa, Iwano et al. 2010). Several recent studies have reported that BPA affects brain development and results in behaviors linked to ASD (Tando, Itoh et al. 2007, Xu, Zhang et al. 2010, Wolstenholme, Edwards et al. 2012, Jardim, Sartori et al. 2017, Kumar and Thakur 2017). ASD children have been reported to exhibit increased levels of BPA in the blood and the urine (Stein, Schluter et al. 2015, Kardas, Bayram et al. 2016, Kondolot, Ozmert et al. 2016). However, whether BPA can alter the expression of genes associated with ASD is unclear. In this study, we therefore sought to determine the association of BPA and ASD at the molecular level by using a series of bioinformatic analyses. Moreover, we predicted biological functions and pathways associated with BPA-responsive genes to determine whether they are associated with ASD-related functions.



## 2. Objectives

1. To predict the association of BPA-responsive genes and ASD using CU-DREAMx bioinformatic software.

2. To predict biological functions and networks of BPA-responsive genes that are related to ASD.

## 3. Methods

### Collection of BPA-responsive genes and ASD candidate genes

To obtain the list of BPA-responsive genes, we searched NCBI GEO Data Sets database (<https://www.ncbi.nlm.nih.gov/gds/>) using “Bisphenol-A” as a search entry to find gene expression studies associated with BPA. All BPA studies which were conducted in cell lines, primary cells, or animals were used in this analysis. ASD candidate genes were collected from SFARI database (<https://gene.sfari.org/database/human-gene/>).

### Association analysis of BPA-responsive genes and ASD candidate genes

The Connection Up and Down Regulation Expression Analysis of Microarrays extension (CU-DREAMx) software is a bioinformatic tool used for identification of differentially expressed genes and for association analysis (Termglinchan, Wanichnopparat et al. 2013). To predict whether BPA can cause

changes in the expression of genes associated with ASD, CU-DREAMx program was used in this study for identifying genes that were significantly disrupted by BPA exposure and also for determining the association between the list of BPA-responsive genes and the list of ASD candidate genes. The gene expression data from each study obtained from NCBI GEO Data Sets was reanalyzed by CU-DREAMx program to identify up- or down-regulated genes in BPA treatment group compared to controls. P-values of less than 0.05 were considered as significantly differentially expressed. The list of significantly differentially expressed genes was then further overlapped with the list of ASD candidate genes obtained from SFARI database using Pearson’s chi-squared test. P-values of less than 0.05 were considered as significantly associated.

### Prediction of biological functions and pathways

Ingenuity Pathway Analysis (IPA) software (<https://www.qiagenbioinformatics.com/products/ingenuity-pathway-analysis/>) was used for predicting biological functions and pathways associated with significantly differentially expressed genes in BPA treatment group that are associated with ASD. Fisher’s exact test (p-value < 0.05) was used for determining significant association.



#### 4. Results

##### **BPA-responsive genes were significantly associated with ASD candidate genes.**

To identify genes that are significantly differentially expressed due to BPA exposure, gene expression data previously deposited in NCBI GEO Data Sets database were downloaded and reanalyzed using CU-DREAMx software as described in the Methods section. Seven transcriptome studies were found to meet the criteria and used for subsequent analyses (**Table 1**). Moreover, the list of up- or down-regulated was then overlapped with the list of 881 ASD candidate genes obtained from autism SFARI databases. Out of seven gene expression studies, the list of BPA-responsive genes from four studies (i.e. GSE44387, GSE63852, GSE50527, and GSE86923) were found to be significantly associated with ASD candidate genes ( $p$ -value $<0.05$ ) (**Table 1**). Interestingly, as

many as 327 genes significantly disrupted in BPA exposure were identified to be ASD candidate genes in SFARI database, suggesting that BPA exposure may cause changes in the genes involved in the molecular mechanisms related to ASD.

##### **Table 1. Association analysis between the list of BPA-responsive genes from each gene expression study and the list of ASD candidate genes identified by SFARI database.**

CU-DREAMx software was employed to identify up- and down-regulated genes that were significantly differentially expressed in BPA treatment group compared to control group in each gene expression dataset. Each list was then overlapped with a total of 881 genes identified to be associated with ASD in SFARI database. Pearson's chi-squared test was conducted to determine the association with ASD candidate genes.



Table 1. Association analysis between the list of BPA-responsive genes from each gene expression study and the list of ASD candidate genes identified by SFARI database.

GEO Datasets	P-values (Chi-Squared Test)	
	Down-Regulated Genes (number of all; number of genes overlapping with SFARI)	Up-Regulated Genes (number of all; number of genes overlapping with SFARI)
GSE5200	6.89E-01 (29; 2)	3.14E-01 (7,851; 424)
GSE44387	6.23E-01 (165; 5)	<b>5.70E-04* (3,889; 114)</b>
GSE63852	7.19E-01 (455; 19)	<b>4.29E-02* (439; 10)</b>
GSE58642	1.31E-01 (332; 7)	3.12E-01 (110; 6)
GSE50527	<b>1.51E-03* (577; 39)</b>	2.79E-01 (684; 34)
GSE58516	4.68E-01 (48; 1)	3.75E-01 (147; 4)
GSE86923	8.09E-02 (2,276; 67)	<b>3.22E-03* (2,986; 89)</b>

#### Biological functions and pathways associated with BPA-responsive genes

To predict biological functions and pathways associated with BPA-responsive genes linked to ASD, the list of 327 BPA-responsive genes identified to be ASD candidate genes were analyzed by Ingenuity Pathway Analysis (IPA) software. The IPA analysis revealed that BPA-responsive genes identified to be ASD candidate genes are significantly associated with “Developmental Disorder” (p-value=5.94E-41-1.42E-04), “Neurological Disease” (p-value=1.24E-33-1.57E-04), and

“Hereditary Disorder” (p-value=6.99E-27-1.42E-04) (Table 2). One of the top canonical pathways significantly associated with this set of BPA-responsive genes is “Reelin Signaling in Neuron” (p-value=2.95E-07) which is a molecular signaling highly implicated in ASD (Table 3). Moreover, the BPA responsive genes were also predicted to significantly associated with “Autism or intellectual disability” (p-value=5.94E-41), “Autism” (p-value=7.33E-14), “Vocalization” (p-value=2.07E-05), and “Social behavior” (p-value=3.06E-05) (Table 4).



**Table 2. Top diseases/disorders significantly associated with BPA-responsive genes.** P-values were calculated by Fisher's exact test (p-value < 0.05).

Diseases/Disorders	P-values	Number of Genes
Developmental Disorder	5.94E-41-1.42E-04	100
Neurological Disease	1.24E-33-1.57E-04	191
Hereditary Disorder	6.99E-27-1.42E-04	127
Organismal Injury and Abnormalities	6.99E-27-1.54E-04	318
Cancer	7.64E-25-1.54E-04	314

**Table 3. Top canonical pathways significantly associated with BPA-responsive genes.** P-values were calculated by Fisher's exact test (p-value < 0.05).

Canonical pathways	P-values
CREB Signaling in Neurons	1.70E-08
GABA Receptor Signaling	3.98E-08
Reelin Signaling in Neurons	2.95E-07
Androgen Signaling	3.47E-07
Synaptic Long Term Depression	1.02E-06



**Table 4. Top neurological diseases/disorders and neurological functions significantly associated with BPA-responsive genes.** P-values were calculated by Fisher's exact test (p-value < 0.05).

Functional Annotation	P-values	Number of Genes
<b>Developmental Disorder</b>		
Autism or intellectual disability	5.94E-41	69
Mental retardation	3.88E-33	59
Familial syndromic intellectual disability	6.24E-21	37
Autism	7.33E-14	18
Susceptibility to autism	2.89E-10	8
<b>Nervous System Development and Function</b>		
Development of central nervous system	3.5E-10	17
Formation of brain	4.54E-08	11
Neuritogenesis	1.03E-06	11
Hearing	7.13E-05	7
<b>Behavior</b>		
Cognition	7.89E-05	8
Vocalization	2.07E-05	4
Social behavior	3.06E-05	4

Gene regulatory network, which is a collection of genes that interact with each other or with other diseases or biological functions, of the BPA-responsive genes were also conducted using IPA software (**Figure 1**). Interestingly, the network shows that BPA-responsive genes are associated with neurological diseases and behaviors related

to ASD including “Autism or intellectual disability”, “Susceptibility to autism type 18”, “Mental retardation”, “Cognitive impairment”, “Hypoplasia of cerebellum” and “Methylation of DNA” (**Figure 1**). Taken together, these findings suggest that BPA exposure may alter the expression of genes involved in biological functions associated with ASD.

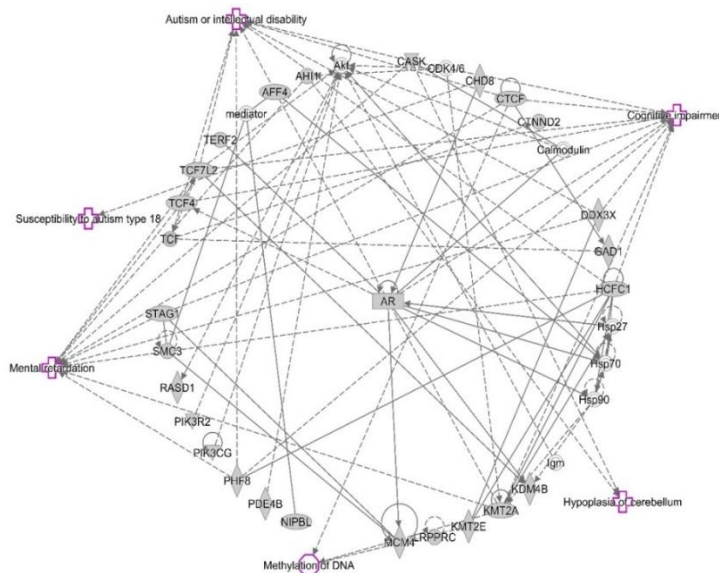


Figure 1. Gene regulatory network of BPA-responsive genes constructed by IPA software.

BPA-responsive genes showed a significant association with dysregulated miRNAs reported in ASD cases.

miRNAs that possibly post-transcriptionally regulate the genes altered by BPA exposure

were also predicted by IPA software. Interestingly, BPA-responsive genes were predicted to be regulated by 65 miRNAs (Table 5).

Table 5. Top miRNAs predicted to be the regulators of BPA-responsive genes.

miRNAs	P-values	BPA-Responsive Target Genes
miR-199a-3p	1.48E-03	<i>MET, MTOR</i>
miR-23a-5p	1.59E-02	<i>LOC102724788/PRODH</i>
miR-29a-5p	1.59E-02	<i>AR</i>
miR-29b-3p	1.07E-02	<i>GPR37, HDAC4, RERE</i>
miR-367-5p	1.59E-02	<i>AR</i>
miR-451a	1.04E-02	<i>BCL2, FBXO33</i>
miR-519a-3p	3.15E-02	<i>AR</i>
miR-93-3p	1.59E-02	<i>AR</i>
mir-133	3.15E-02	<i>RB1CC1</i>
mir-185	6.62E-03	<i>AR, EPHB2</i>



### Overlapping of BPA-responsive genes showed candidate genes for further ASD studies

To find BPA-responsive genes overlapping among gene expression studies, we conducted the overlap analysis using the Venn Diagram software (<http://bioinformatics.psb.ugent.be/webtools/Venn/>) (Figure 2). It is noticeable that BPA-responsive genes repeatedly found to be dysregulated by BPA exposure in at least three independent studies are *SOD1*, *MEF2C*, and *GNAS*. These genes may serve as good candidates for further studies.

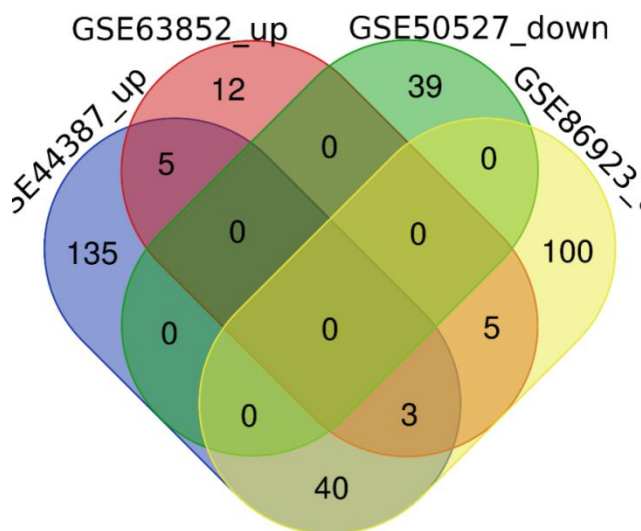


Figure 2. Venn diagram of BPA-responsive genes that are related to ASD.

### 5. Discussion

This is the first study to demonstrate that exposure to BPA may alter the expression profiles of genes linked to ASD. Using CU-DREAMx bioinformatic analysis of gene expression data from BPA studies in NCBI GEO DataSets database and ASD candidate genes, we found that BPA-responsive genes from four studies were significantly associated with ASD candidate genes. Interestingly, the dysregulated genes found to be reproducibly identified in at least

three independent studies are *SOD1*, *MEF2C*, and *GNAS*. *SOD1* (superoxide dismutase 1) encodes the protein that is responsible for destroying free superoxide radicals in the body. Oxidative stress is found to be associated with ASD etiology and/or susceptibility see reviews in (Kern and Jones 2006, Frustaci, Neri et al. 2012, Rossignol and Frye 2014). A recent study have identified single nucleotide polymorphisms (SNPs) in *SOD1* genes in children with ASD (Kovac, Macedoni Luksic et



al. 2014). *MEF2C* (Myocyte Enhancer Factor 2C) encodes a transcription factor involved in diverse developmental processes, including neurogenesis. Deletion in *MEF2C* has been reported in individuals with ASD (Novara, Beri et al. 2010). *GNAS* encodes a protein that functions as transducers in numerous signaling pathways controlled by G protein-coupled receptors. SNPs and mutation in *GNAS* have been identified in individuals with ASD (Sanders, Murtha et al. 2012). These results suggest that BPA exposure may increase the risk of ASD by causing the dysregulation of these ASD-related genes.

Next, IPA analysis revealed that the list of BPA-responsive genes were associated with “**Reelin signaling pathway**”. Reelin protein was found to be impaired in ASD (Zhang, Liu et al. 2002), suggesting that BPA exposure may cause aberrant gene expression profiles involved in pathobiology in ASD. Our gene regulatory network predicted by IPA showed a significant association between the BPA-responsive genes and biological functions related to ASD which are “Autism or intellectual disability”, “Susceptibility to autism type 18”, “Mental retardation”, “Cognitive impairment”, “Hypoplasia of cerebellum” and “Methylation of DNA”. The cerebellum is known to have functions in motor learning and coordination. Recent studies have reported that ASD cases

exhibited the loss of Purkinje cells in the cerebellum (Bailey, Luthert et al. 1998, Palmen, van Engeland et al. 2004). Moreover, cerebellar hypoplasia which is a condition which the patients have a smaller size of cerebellum is also associated with ASD (Courchesne, Yeung-Courchesne et al. 1988). Altered methylation was also significantly associated with ASD cases (Nguyen, Rauch et al. 2010, Saeli, Tangsuwansri et al. 2018). Moreover, the prediction of miRNAs regulating these BPA-responsive ASD candidate genes revealed as many as 65 miRNAs. Interestingly, a total 13 miRNAs has been reported in ASD. These miRNAs include miR-23a, miR-367, miR-93, and miR-185 which has been reported to be significantly down-regulated in ASD and miR-133 which was significantly up-regulated (Sarachana, Zhou et al. 2010). These results suggest that BPA exposure may also impair the expression of miRNAs which regulate the expression of several ASD-related genes. The molecular mechanisms through which BPA exerts its effect and impact ASD-related gene expression deserve further investigation.

## 6. Conclusion

The findings from this study suggest that BPA exposure may lead to changes in the expression of genes and miRNAs associated



with biological functions and pathways associated with ASD. BPA-responsive genes from several transcriptomic studies were significantly associated with ASD candidate genes. Prediction of biological functions associated with BPA-responsive ASD candidate genes revealed ASD-related canonical pathways and neurological functions and disorders including “Reelin signaling pathway”, “autism or intellectual disability”, which are significantly associated with ASD. The role of BPA exposure in the etiology and susceptibility of ASD should be studied further.

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## 8. References

1. Adams, J. B., T. Audhya, S. McDonough-Means, R. A. Rubin, D. Quig, E. Geis, E. Gehn, M. Loresto, J. Mitchell, S. Atwood, S. Barnhouse and W. Lee (2013). "Toxicological status of children with autism vs. neurotypical children and the association with autism severity." *Biol Trace Elem Res* **151**(2): 171-180.
2. Bailey, A., P. Luthert, A. Dean, B. Harding, I. Janota, M. Montgomery, M. Rutter and P. Lantos (1998). "A clinicopathological study of autism." *Brain* **121 ( Pt 5)**: 889-905.
3. Baio, J., L. Wiggins, D. L. Christensen, M. J. Maenner, J. Daniels, Z. Warren, M. Kurzius-Spencer, W. Zahorodny, C. Robinson Rosenberg, T. White, M. S. Durkin, P. Imm, L. Nikolaou, M. Yeargin-Allsopp, L. C. Lee, R. Harrington, M. Lopez, R. T. Fitzgerald, A. Hewitt, S. Pettygrove, J. N. Constantino, A. Vehorn,



- J. Shenouda, J. Hall-Lande, K. Van Naarden Braun and N. F. Dowling (2018). **"Prevalence of Autism Spectrum Disorder Among Children Aged 8 Years - Autism and Developmental Disabilities Monitoring Network, 11 Sites, United States, 2014."** MMWR Surveill Summ **67**(6): 1-23.
4. Courchesne, E., R. Yeung-Courchesne, G. A. Press, J. R. Hesselink and T. L. Jernigan (1988). **Hypoplasia of cerebellar vermal lobules VI and VII in autism."** N Engl J Med **318**(21): 1349-1354.
5. Frustaci, A., M. Neri, A. Cesario, J. B. Adams, E. Domenici, B. Dalla Bernardina and S. Bonassi (2012). **"Oxidative stress-related biomarkers in autism: systematic review and meta-analyses."** Free Radic Biol Med **52**(10): 2128-2141.
6. Geier, D. A., T. Audhya, J. K. Kern and M. R. Geier (2010). **"Blood mercury levels in autism spectrum disorder: Is there a threshold level?"** Acta Neurobiol Exp (Wars) **70**( 2): 177-186.
7. Jardim, N. S., G. Sartori, M. H. M. Sari, S. G. Muller and C. W. Nogueira (2017). **"Bisphenol A impairs the memory function and glutamatergic homeostasis in a sex-dependent manner in mice: Beneficial effects of diphenyl diselenide."** Toxicol Appl Pharmacol **329**: 75-84.
8. Kardas, F., A. K. Bayram, E. Demirci, L. Akin, S. Ozmen, M. Kendirci, M. Canpolat, D. B. Oztop, F. Narin, H. Gumus, S. Kumandas and H. Per (2016). **"Increased Serum Phthalates (MEHP, DEHP) and Bisphenol A Concentrations in Children With Autism Spectrum Disorder: The Role of Endocrine Disruptors in Autism Etiopathogenesis."** J Child Neuro **31**(5): 629-635.
9. Kern, J. K. and A. M. Jones (2006). **"Evidence of toxicity, oxidative stress, and neuronal insult in autism."** J Toxicol Environ Health B Crit Rev **9**(6): 485-499.
10. Kondolot, M., E. N. Ozmert, A. Asci, P. Erkekoglu, D. B. Oztop, H. Gumus, B. Kocer-Gumusel and K. Yurdakok (2016). **"Plasma phthalate and bisphenol a levels and oxidant-antioxidant status in autistic children."** Environ Toxicol Pharmacol **43**: 149-158.
11. Kovac, J., M. Macedoni Luksic, K. Trebusak Podkrajsek, G. Klancar and T. Battelino (2014). **"Rare single nucleotide polymorphisms in the regulatory regions of the superoxide dismutase**



- genes in autism spectrum disorder." Autism Res 7(1):138-144.
12. Kumar, D. and M. K. Thakur (2017). "Anxiety like behavior due to perinatal exposure to Bisphenol-A is associated with decrease in excitatory to inhibitory synaptic density of male mouse brain." Toxicology 378: 107-113.
13. LaSalle, J. M. (2013). "Autism genes keep turning up chromatin." OA Autism 1(2): 14-.
14. Miodovnik, A., S. M. Engel, C. Zhu, X. Ye, L. V. Soorya, M. J. Silva, A. M. Calafat and M. S. Wolff (2011). "Endocrine disruptors and childhood social impairment." Neurotoxicology 32(2): 261-267.
15. Moosa, A., H. Shu, T. Sarachana and V. W. Hu (2018). "Are endocrine disrupting compounds environmental risk factors for autism spectrum disorder?" Horm Behav 101: 13-21.
16. Nguyen, A., T. A. Rauch, G. P. Pfeifer and V. W. Hu (2010). "Global methylation profiling of lymphoblastoid cell lines reveals epigenetic contributions to autism spectrum disorders and a novel autism candidate gene, RORA, product is reduced in autistic brain." Fasebj 24(8): 3036-3051.
17. Nishikawa, M., H. Iwano, R. Yanagisawa, N. Koike, H. Inoue and H. Yokota (2010). Placental transfer of conjugated bisphenol A and subsequent reactivation in the rat fetus." Environ Health Perspect 118(9): 1196-1203.
18. Novara, F., S. Beri, R. Giorda, E. Ortibus, S. Nageshappa, F. Darra, B. Dalla Bernardina, O. Zuffardi and H. Van Esch (2010). "Refining the phenotype associated with MEF2C haploinsufficiency." Clin Genet 78(5): 471-477.
19. Palmen, S. J., H. van Engeland, P. R. Hof and C. Schmitz (2004). "Neuropathological findings in autism." Brain 127(Pt 12): 2572-2583.
20. Roberts, E. M., P. B. English, J. K. Grether, G. C. Windham, L. Somberg and C. Wolff (2007). "Maternal residence near agricultural pesticide applications and autism spectrum disorders among children in the California Central Valley." Environ Health Perspect 115(10): 1482-1489.
21. Rossignol, D. A. and R. E. Frye (2014). "Evidence linking oxidative stress,



- mitochondrial dysfunction, and inflammation in the brain of individuals with autism." *Front Physiol* 5: 150.
22. Saeliw, T., C. Tangsuwansri, S. Thongkorn, W. Chonchaiya, K. Suphapeetiporn, A. Mutirangura, T. Tencomnao, V. W. Hu and T. Sarachana (2018). "Integrated genome-wide Alu methylation and transcriptome profiling analyses reveal novel epigenetic regulatory networks associated with autism spectrum disorder." *9*: 27.
23. Sanders, S. J., M. T. Murtha, A. R. Gupta, J. D. Murdoch, M. J. Raubeson, A. J. Willsey, A. G. Ercan-Sencicek, N. M. DiLullo, N. N. Parikshak, J. L. Stein, M. F. Walker, G. T. Ober, N. A. Teran, Y. Song, P. El-Fishawy, R. C. Murtha, M. Choi, J. D. Overton, R. D. Bjornson, N. J. Carriero, K. A. Meyer, K. Bilguvar, S. M. Mane, N. Sestan, R. P. Lifton, M. Gunel, K. Roeder, D. H. Geschwind, B. Devlin and M. W. State (2012). "De novo mutations revealed by whole-exome sequencing are strongly associated with autism." *Nature* 485(7397): 237-241.
24. Sarachana, T., R. Zhou, G. Chen, H. K. Manji and V. W. Hu (2010). "Investigation of post-transcriptional gene regulatory networks associated with autism spectrum disorders by microRNA expression profiling of lymphoblastoid cell lines." *Genome Med* 2(4): 23.
25. Stein, T. P., M. D. Schluter, R. A. Steer, L. Guo and X. Ming (2015). "Bisphenol Exposure in Children With Autism Spectrum Disorders." *Autism Res* 8(3): 272-283.
26. Sun, W., J. Poschmann, R. Cruz-Herrera Del Rosario, N. N. Parikshak, H. S. Hajan, V. Kumar, R. Ramasamy, T. G. Belgard, B. Elanggovan, C. C. Y. Wong, J. Mill, D. H. Geschwind and S. Prabhakar (2016). "Histone Acetylome-wide Association Study of Autism Spectrum Disorder." *Cell* 167(5): 1385-1397.e1311.
27. Sun, Y., M. N. Nakashima, M. Takahashi, N. Kuroda and K. Nakashima (2002). "Determination of bisphenol A in rat brain by microdialysis and column switching high-performance liquid chromatography with fluorescence detection." *Biomed Chromatogr* 16(5): 319-326.
28. Tando, S., K. Itoh, T. Yaoi, J. Ikeda, Y. Fujiwara and S. Fushiki (2007). "Effects of pre-and neonatal exposure to bisphenol A on *Dev* 29(6): 352 - 356.



29. Termglinchan,V., W. Wanichnopparat, K. Suwanwongse, C. Teeyapant, K. Chatperporn, K. Leerunyakul, K. Chuadpia, O. Sirimaneethum, P. Wijitworawong, W. Mutirangura, C. Apornthewan, C. Vinayanuwattikun and A. Mutirangura (2013). **"Candidate cancer-targeting agents identified by expression-profiling arrays."** Onco Targets Ther **6**: 447-458.
30. Wolstenholme, J. T., M. Edwards, S. R. Shetty, J. D. Gatewood, J. A. Taylor, E. F. Rissman and J. J. Connelly (2012). **"Gestational exposure to bisphenol a produces transgenerational changes in behaviors and gene expression."** Endocrinology **153**(8): 3828-3838.
31. Xu, X. H., J. Zhang, Y. M. Wang, Y. P. Ye and Q. Q. Luo (2010). **"Perinatal exposure to bisphenol-A impairs learning-memory by concomitant down-regulation of N - methyl-D-aspartate receptors of hippocampus in male offspring mice."** Horm Behav **58**(2): 326-333.
32. Zhang, H., X. Liu, C. Zhang, E. Mundo, F. Macciardi, D. R. Grayson, A. R. Guidotti and J. J. Holden (2002). **"Reelin gene alleles and susceptibility to autism spectrum disorders."** Mol Psychiatry**7**(9): 1012-1017.